

REVIEW ARTICLE

GOT2: New therapeutic target in pancreatic cancer



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Abstract In recent years, the incidence and mortality rates of pancreatic cancer have been steadily increasing, and conventional therapies have shown a high degree of tolerance. Therefore, the search for new therapeutic targets remains a key issue in current research. Mitochondrial glutamic-oxaloacetic transaminase 2 (GOT2) is an important component of the malate-aspartate shuttle system, which plays an important role in the maintenance of cellular redox balance and amino acid metabolism, and has the potential to become a promising target for anti-cancer therapy. In this paper, we will elaborate on the metabolic and immune effects of GOT2 in pancreatic cancer based on existing studies, with a view to opening up new avenues for the treatment of pancreatic cancer.

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Introduction

Pancreatic cancer is a common malignant tumor in the digestive system. Existing data indicate that the incidence of pancreatic cancer has been increasing in recent years, while its 5-year overall survival rate has been decreasing, and it is expected to become the second leading cause of cancer-related death by 2030.^{1,2} In terms of treatment, due to its insensitivity to most chemotherapeutic agents and

poor therapeutic response, surgical resection is currently the only curative method for pancreatic cancer. Unfortunately, pancreatic cancer has an insidious onset and inconspicuous clinical symptoms. 80%–85% of patients are diagnosed at an advanced stage that is difficult to resect, and there is a high recurrence rate even after surgical treatment.³ With the continuous deepening of research in recent years, metabolic therapy and immunotherapy have gradually entered people's vision. The research on metabolic or immune targets of pancreatic cancer may provide new ideas for its clinical treatment.

Glutamic-oxaloacetic transaminases (GOTs), also known as aspartate transaminases, play a crucial role in amino acid metabolism and the tricarboxylic acid (TCA) cycle and can be classified into two subtypes according to their sub-cellular localization, namely GOT1 in the cytoplasm and

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GOT2 in the mitochondria. In normal physiology, GOTs play key roles in cellular energy metabolism. Firstly, GOTs participate in glutamine metabolism and mediate the interconversion of glutamate and oxaloacetate with α -ketoglutarate and aspartate by catalyzing transamination reaction, the products of which directly promote cell proliferation. For example, aspartate (Asp) serves as a precursor for the synthesis of many amino acids and provides raw materials for protein and nucleotide synthesis,^{4,5} while α -ketoglutarate (α -KG) plays a critical role in the production of ATP and the replenishment of intermediates in the TCA cycle.⁶ Secondly, GOTs are key components of the malate-aspartate shuttle system,⁷ which enable NADH to transfer between the cytoplasm and mitochondria, and maintain cellular redox balance. Recent studies have revealed that apart from its traditional metabolic functions, there is also an unconventional function of GOT2 in the nucleus, which may be achieved through fatty acid

binding and uptake.⁸ Based on the above key roles of GOT2 in cells, researchers have deduced that GOT2 may be a potential therapeutic target for various cancers and have initiated relevant studies.

In this paper, we will comprehensively and systematically elaborate on the metabolic and immune functions of GOT2 and the corresponding molecular mechanisms in pancreatic cancer based on existing research (Fig. 1), with the aim of providing new ideas and approaches for pancreatic cancer treatment.

Metabolic roles of GOT2 in pancreatic cancer

The occurrence and development of tumors depend on cellular metabolic reprogramming.^{9,10} Metabolic therapy focuses on the metabolic dependence of cancer cells.¹¹ Therefore, it is essential to fully understand how

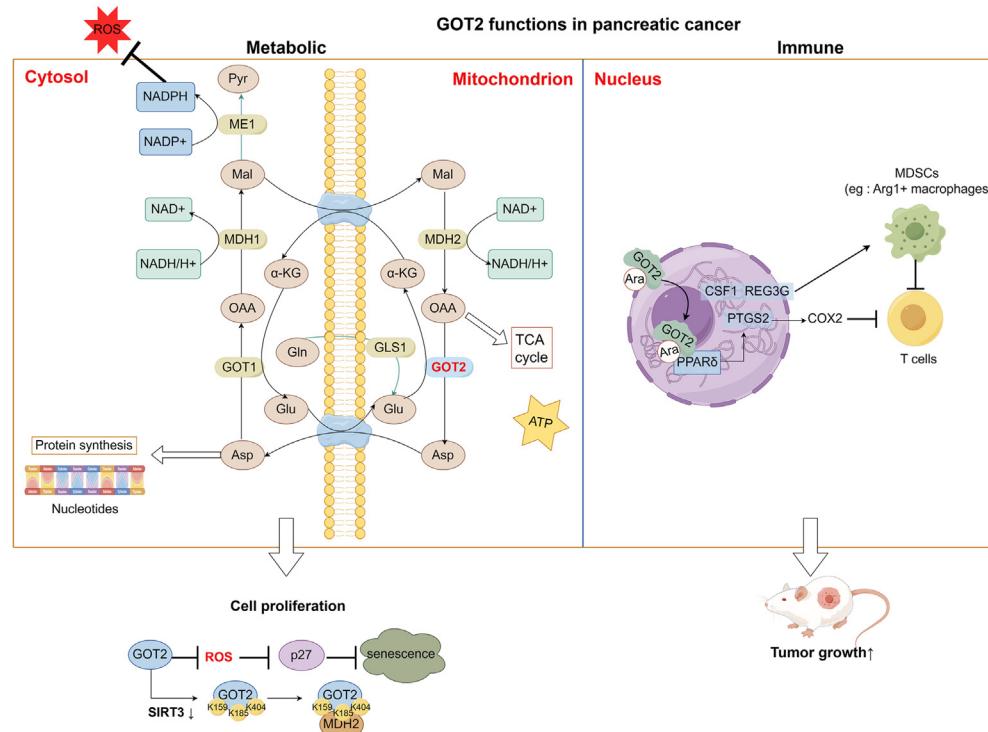


Figure 1 Metabolic and immune functions of GOT2 in pancreatic cancer cells. First, GOT2 catalyzes transamination reaction in mitochondria, and the products can directly promote cell proliferation. Aspartate provides raw materials for protein and nucleotide synthesis, and oxaloacetate participates in the TCA cycle. Second, GOT2 is a key component of the malate-aspartate shuttle system, which maintains cellular redox balance. In addition, pancreatic cancer cells exhibit unique glutamine metabolism. On this basis, decreased SIRT3 expression leads to an increased level of GOT2 acetylation. The acetylation of GOT2 at 3K sites enhances the protein association of GOT2-MDH2, which promotes the malate-aspartate NADH shuttle activity to increase ATP production and stimulates NADPH production to inhibit ROS, thereby reducing the expression of cyclin-dependent kinase inhibitor p27 and suppressing cellular senescence (leftmost plot). An unexpected nuclear role of GOT2 in pancreatic cancer: As a nuclear fatty acid transporter, GOT2 promotes the transcriptional activity of PPAR δ by directly binding to fatty acids (mainly arachidonic acid) and induces the expression of immunomodulatory target genes. PTGS2 encodes COX2 that inhibits T cell-mediated anti-tumor immunity. CSF1 and REG3G encodes secretory protein which recruits myeloid-derived suppressor cells (MDSCs) into the tumor microenvironment (right plot). OAA, oxaloacetate; α -KG, α -ketoglutarate; Ara, arachidonic acid; Asp, aspartate; COX2, cyclooxygenase 2; CSF1, colony stimulating factor 1; Glu, glutamine; Gln, glutamine; GOT2, glutamic-oxaloacetic transaminase 2; Mal, malate; MDH2, malate dehydrogenase 2; PPAR δ , peroxisome proliferator-activated receptor δ ; PTGS2, prostaglandin endoperoxide synthase 2; Pyr, pyruvate; REG3G, regenerating islet-derived protein 3 gamma; ROS, reactive oxygen species; SIRT3, sirtuin-3; TCA, tricarboxylic acid. This picture was drawn by Figdraw.

metabolism is regulated in tumor cells. Pancreatic cancer cells are strongly dependent on unique glutamine metabolism, which utilizes glutamine (Gln)-derived Asp to maintain cell redox state.¹² Targeting specific metabolic enzymes in this pathway may bring new hope for pancreatic cancer treatment.

Enzymes involved in the glutamine metabolic reprogramming (mitochondrial GLS1 (glutaminase 1), GOT1, and GOT2) are highly up-regulated in pancreatic cancer.¹³ In mitochondria, Gln is catabolized to glutamate via GLS1 and further converted into TCA cycle intermediates α -KG and Asp via GOT2, providing raw materials for mitochondrial energy metabolism. Next, Asp is transported to the cytoplasm and converted to oxaloacetate via GOT1, and then to malate, which is finally catalyzed by malic enzyme 1 (ME1) to generate pyruvate and produce NADPH to inhibit reactive oxygen species (ROS) generated during rapid cell proliferation and maintain cellular redox homeostasis.¹² This metabolic reprogramming of pancreatic cancer is mainly mediated by mutant KRAS (KRAS*), and that oncogenic KRAS* is the signature genetic event for pancreatic cancer progression.^{14,15} Non-canonical glutamine metabolism is crucial for pancreatic cancer growth. Targeting this pathway has become a new approach for pancreatic cancer treatment.^{16–18}

GOT2 plays a critical role as a component of glutamine metabolic reprogramming. Currently, studies have demonstrated that GOT2 depletion leads to impaired proliferation and disrupted redox homeostasis in pancreatic cancer cells *in vitro*.¹⁹ Consistent with its conventional metabolic effects, GOT2 knockdown (KD) in cultured pancreatic cancer cells results in decreased Asp and α -KG production, thereby preventing the normal TCA cycle and decreasing ATP levels. This suggests that GOT2 promotes glutamine anaplerosis. Second, GOT2 KD forces the malate-aspartate shuttle to interrupt and prevents the transfer of NADH from the cytoplasm to the mitochondria, which induces cellular NADH accumulation and reductive stress, and results in the block of glyceraldehyde 3-phosphate dehydrogenase (GADPH) node and impaired glycolytic function. Meanwhile, loss of GOT2 leads to a decrease in cellular NADPH/NADP⁺ ratio and a significant increase in ROS levels,²⁰ which could be aggravated under hypoxic conditions.²¹

Existing studies have explored this pathway to regulate redox homeostasis to obtain more effective treatment strategies for pancreatic cancer. GOT2 can inhibit pancreatic cancer cell senescence by maintaining cellular redox balance.²⁰ Here, GOT2 KD induces the expression of the cyclin-dependent kinase inhibitor p27 by increasing ROS levels in the cells and triggers its mediated cellular senescence. Relevant literature suggests that p27 is down-regulated in several human cancers and plays a critical role in regulating cell cycle progression and inducing and maintaining senescence.^{22,23} Since KRAS* and combinations of mutation in p53 and p16²⁴ may allow pancreatic cancer cells to bypass classical senescence pathways, targeting senescence pathways regulated by GOT2 may be important in overcoming therapeutic resistance in pancreatic cancer. In addition, SIRT3 (sirtuin-3)-dependent GOT2 acetylation stimulates the malate-aspartate NADH shuttle activity and oxidative protection, thereby promoting pancreatic tumor

growth.²⁵ GOT2 acetylation at lysine residues K159, K185, and K404 (3K), enhances the protein association between GOT2 and malate dehydrogenase 2 (MDH2), thereby facilitating malate-aspartate shuttle activity. SIRT3 has been identified as the main deacetylase of GOT2, and its deacetylation may not affect the enzyme activity of GOT2 itself but impairs the GOT2-MDH2 association, which negatively regulates malate-aspartate shuttle and impairs pancreatic cancer cell proliferation. Thus, SIRT3 may act as a tumor suppressor in pancreatic cancer,²⁶ and GOT2 deacetylation drugs targeting this need to be explored. Given the elevated levels of GOT2 K159 acetylation in pancreatic cancer, this could be a potential biomarker for pancreatic cancer diagnosis.²⁵

Notably, GOT2 is essential for maintaining growth and inhibiting senescence in pancreatic cancer, but has little effect on proliferation and senescence in untransformed human pancreatic ductal cells, which may be related to the greater dependence of normal cells on glutamate dehydrogenase 1 (GLUD1).^{12,20} Therefore, targeting GOT2 may provide sufficient therapeutic window to protect the surrounding tissues from damage and become one of the effective and specific strategies for treating pancreatic cancer. Impaired Gln metabolism has been found to cause pancreatic cancer cells to be more sensitive to oxidative stress,¹² which may have synergy with treatments that increase cellular ROS levels, such as radiotherapy and chemotherapy. In addition, recently developed targeted drugs, such as KRAS* selective inhibitors^{27,28} and GLS inhibitors,^{13,29,30} are in various stages of preclinical or clinical trials and are considered to have important value in pancreatic cancer treatment, but still have limitations such as resistance or weak efficacy of single therapy. So GOT2-targeted therapy may be an ideal combination or alternative approach and is expected to have a profound impact on future treatment.

Overall, GOT2 as a hub for glutamine metabolic reprogramming contributes to cancer progression by providing Asp and α -KG for biosynthesis and energy requirements. In fact, various cancers show sensitivity to glutamine deprivation.¹⁶ However, GOT2 has different regulatory mechanisms in different tumors,^{31–35} giving us a new perspective on how GOT2-mediated glutamine metabolism functions in cancers. BRCA1/ZBRK1 repressor complex reduces Asp biosynthesis through transcriptional repression of GOT2 expression, thereby leading to impaired proliferation of breast cancer cells.³¹ In non-small cell lung cancer, miRNA down-regulates GOT2 at post-transcriptional level. Silencing circ-SEC31A significantly affects malate-aspartate metabolism, and inhibits cell proliferation, migration, and invasion by regulating the miR-520a-5p/GOT2 axis.³² Importantly, although hepatocellular carcinoma (HCC) similarly exhibits glutamine addiction, GOT2 is lowly expressed in HCC cells, which is related to advanced progression and poor prognosis.³³ Here, GOT2 KD maintains glutathione/ROS balance by facilitating glutathione synthesis, thereby activating the PI3K/AKT/mTOR pathway and ultimately promoting HCC progression. GOT2 plays a suppressive and promoting role in HCC and pancreatic cancer, respectively, which may be associated with generating glutathione through different pathways to maintain cellular redox balance.

Immune role of GOT2 in pancreatic cancer

Recently, a study has revealed an unexpected nuclear role of GOT2 in pancreatic cancer. Distinct from its traditional metabolic function, GOT2 acts as a nuclear fatty acid transporter to promote the transcriptional activity of peroxisome proliferator-activated receptor δ (PPAR δ) by directly binding to fatty acids and inhibit T cell-mediated anti-tumor immunity, thereby facilitating pancreatic cancer progression.⁸

Abrego et al first assessed the significance of GOT2 on pancreatic cancer progression and found GOT2 knockdown had little or no impact on the proliferation of pancreatic cancer cells *in vitro*. Similarly, in immunodeficient mouse transplantation models, tumor growth did not change after silencing GOT2. However, tumor growth was severely impaired in immune-functioning syngeneic mice. Consistent with the results *in vitro*, tumor cell proliferation *in vivo* was not impaired. These results suggest that the effect of GOT2 on tumor growth *in vivo* may be immune-mediated. In accordance with this view, GOT2-silenced tumor cells showed an increase in T-cell content, including CD4 $^{+}$ and CD8 $^{+}$ T cells, and a decrease in immunosuppressive Arg1 $^{+}$ macrophage abundance. After treatment with neutralizing antibodies against T cells, GOT2-silenced tumor growth was restored. The authors further found that GOT2 was localized in the nucleus in pancreatic cancer cells and acted as a fatty acid-binding protein to regulate the transport of nuclear fatty acids, and bind to and activate PPAR δ , promoting its transcriptional activity. PPAR δ undergoes conformational changes after binding to nuclear fatty acids, altering its binding to DNA and inducing the expression of target genes.³⁶ Prostaglandin endoperoxide synthase 2 (PTGS2) is one of the immunomodulatory genes regulated by PPAR δ and encodes COX2 (cyclooxygenase 2) which induces prostaglandin synthesis. Recent reports have revealed that COX2 contributes to the reduction of T-cell infiltration in the pancreatic cancer microenvironment³⁷ and is associated with anti-tumor immunosuppression in other cancers.³⁸ In addition, other genes activated by PPAR δ , such as CSF1 (colony stimulating factor 1) and REG3G (regenerating islet-derived protein 3 gamma), encode secreted proteins that recruit myeloid-derived suppressor cells to the tumor microenvironment. Furthermore, arachidonic acid has been identified as the major GOT2-binding fatty acid mediating the GOT2-PPAR δ interaction.

The study of Abrego and colleagues has revealed the part-time function of the metabolic enzyme GOT2 in pancreatic cancer: the GOT2-PPAR δ axis significantly regulates the immune microenvironment of pancreatic cancer to inhibit anti-tumor immunity. This provides new insights into pancreatic cancer treatment and prompts us to consider whether targeting GOT2 to promote immune response could extend beyond pancreatic cancer cells. Recent studies have revealed the direct roles of GOT2 in immune cells.³⁹⁻⁴¹ In preclinical models, co-expression of exogenous GOT2 enhances the metabolism and proliferative capacity of anti-GPC3 CAR-T cells and significantly increases the anti-tumor activity of CAR-T cells in specific solid tumors.³⁹ The importance of GOT2 in T cells is due to its metabolic roles. Another study showed that GOT2 plays

an important role in the metabolic reprogramming of diffuse large B-cell lymphoma.⁴⁰ GOT2 provides Asp and nucleotides to cells with activated or abnormal Jak/STAT and NF- κ B signaling pathways, thereby promoting cell proliferation. In addition, GOT2 and GLS as prognostic biomarkers for breast cancer, are closely associated with dendritic cells and immunotherapy response.⁴¹ Cluster 2 (lower GOT2 expression and higher GLS expression) shows a better immunotherapy response. In short, these findings suggest the possibility of establishing a close link between glutamine metabolism and immunity for cancer treatment, which allows us to overcome tumor immune evasion and adapt immunotherapies to improve patient outcomes.

Alternative mechanisms by which pancreatic cancer cells bypass GOT2 silencing

Pancreatic cancer exhibits a unique tumor microenvironment.^{42,43} Due to the poor vascular function of pancreatic tumors, pancreatic cancer cells are exposed to a long-term hypoxic microenvironment and severely restricted in terms of nutrients,⁴⁴ which damages the ability to transfer electrons to oxygen in the mitochondria, thereby limiting Asp synthesis.^{4,5} Under hypoxic conditions, pancreatic cancer cells depend on GOT2 to synthesize endogenous Asp.^{21,45} However, pancreatic tumors *in vivo* may use alternative pathways to obtain Asp, thereby bypassing the dependence on GOT2. Two recent studies have elucidated the compensatory mechanisms of pancreatic cancer cells to overcome GOT2 loss *in vivo*, based on cell extrinsic and cell intrinsic factors (Fig. 2).^{19,21}

Kerk et al found that GOT2 deficiency impaired pancreatic cancer cell proliferation *in vitro* but had no effect on tumor growth *in vivo*.¹⁹ Given that pancreatic cancer exhibits a unique GOT2-dependent glutamine metabolism, there must be other compensatory metabolic pathways for pancreatic cancer cells *in vivo* to survive under GOT2-deficient conditions. Cancer-associated fibroblasts (CAFs) are major components of the pancreatic tumor microenvironment,⁴⁶ such as human pancreatic stellate cells. CAFs indirectly compensate for the insufficient Asp due to GOT2 deficiency by releasing the redox-active metabolite pyruvate to GOT2-deficient pancreatic cancer cells. Here, pyruvate, as an electron acceptor, is transported into pancreatic cancer cells via monocarboxylate transporter 1 (MCT1) and converted to lactate via lactate dehydrogenase (LDH), which can restore the redox imbalance caused by GOT2 deficiency and reverse the GOT1 pathway to synthesize Asp. CAFs conditioned medium can rescue the proliferation of GOT2-deficient pancreatic cancer cells *in vitro*, while inhibiting pyruvate uptake and metabolism can block their recovery. However, these interventions are ineffective against GOT2 KD tumors *in vivo*, which may indicate the presence of other Asp sources in pancreatic tumor microenvironment.

Macropinocytosis is a lysosomal degradation pathway driven by pancreatic cancer cells expressing KRAS*, which can provide enough Asp to support pancreatic cancer cell to survive in nutritional constraints by capturing environmental albumin and releasing amino acids within the

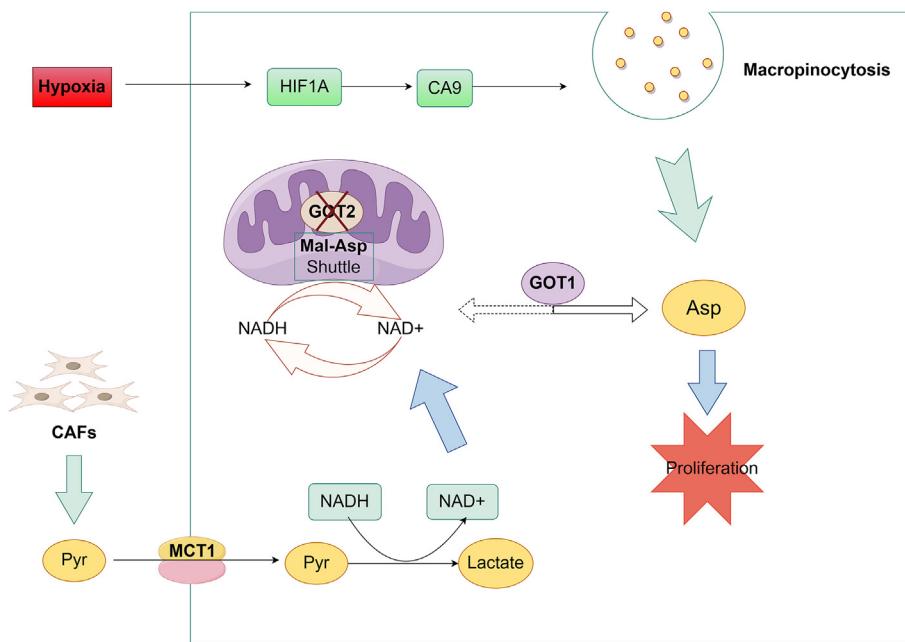


Figure 2 Alternative mechanisms bypass GOT2 silencing in pancreatic cancer cells. CAFs can secrete pyruvate in the pancreatic tumor microenvironment. Here pyruvate is transported to pancreatic cancer cells via MCT1 and converted into lactate, which can restore the redox imbalance caused by GOT2 loss and reverse the GOT1 pathway to synthesize Asp, thus saving pancreatic cancer cell proliferation *in vitro*. Under hypoxia conditions, HIF1A enhances the effect of macropinocytosis by inducing CA9 expression. Macropinocytosis can directly provide enough Asp to GOT2-deficient pancreatic cancer cells by capturing environmental albumin and releasing amino acids inside the cells. These two alternative routes for acquiring Asp allow pancreatic cancer to bypass the metabolic limitations from GOT2 loss. Asp, aspartate; CA9, carbonic anhydrase IX; CAFs, cancer-associated fibroblasts; GOT1/2, glutamic-oxaloacetic transaminase 1/2; HIF1A, hypoxia-inducible factor 1 subunit α ; MCT1, monocarboxylate transporter 1; Pyr, pyruvate. This picture was drawn by Figdraw.

cells.^{47–49} Thus, this nutrient scavenging pathway leads to pancreatic cancer resistance to therapies targeting non-classical metabolic pathways. Garcia-Bermudez et al revealed a relationship between hypoxia and macropinocytosis.²¹ Macropinocytosis supports the proliferation of hypoxic pancreatic cancer cells, and hypoxia in turn enhances the effect of macropinocytosis. Here, hypoxia-inducible factor 1 subunit α (HIF1A) induces carbonic anhydrase IX (CA9) expression⁵⁰ and increases the production of its catalytic product bicarbonate, thereby up-regulating macropinocytosis. Currently, inhibitors targeting scavenging pathway have made progress in preclinical studies or clinical trials in pancreatic cancer, including lysosome acidification inhibitors^{51,52} and CA9 inhibitors.⁵³ New and more potent autophagy inhibitors are also being developed,^{54,55} which may block pancreatic cancer cells from acquiring Asp via macropinocytosis and hopefully slow the growth of GOT2 KD pancreatic tumors.

The above two studies suggest that pancreatic cancer cells interact with the tumor microenvironment to open up alternative routes to acquire Asp, allowing pancreatic cancer to bypass the metabolic constraints from GOT2 deficiency. In the complex tumor microenvironment of pancreatic cancer, in addition to the metabolism of pancreatic cancer cells themselves, the metabolism between stromal cells and immune cells can also regulate tumor progression.¹⁰ Therefore, it is important to consider the role of tumor microenvironment in metabolic therapies

targeting GOT2 in pancreatic cancer and gain a better understanding of its potential resistance mechanisms, which will be beneficial to develop more effective metabolic therapies.

Conclusion

According to existing research, we have a deeper understanding of the roles of GOT2 in pancreatic cancer, and in addition to its well-known metabolic significance, the unexpected role of immunosuppression also deserves our attention. Cellular metabolic reprogramming and intrinsic immune escape are key features of pancreatic cancer, and GOT2, as a metabolic enzyme, bridges organically the two, presenting a broader prospect for targeting GOT2 to treat pancreatic cancer.

Small molecule inhibitors of GOT2 are expected to be effective therapeutics for pancreatic cancer. Studies have shown that the pan-transaminase inhibitor amino oxy-acetate (AOA) is a promising strategy for GOT2-targeted cancer therapy. By inhibiting malate-aspartate shuttle, AOA can damage the pathway of glucose conversion to TCA cycle products, thereby inhibiting the proliferation of breast cancer cells *in vitro* and tumor growth in xenograft models of athymic mice.⁵⁶ AOA treatment selectively induced the death of glutamine-dependent MYC-amplified glioblastoma cell line *in vitro* without affecting the

viability of Myc-deficient cell line.⁵⁷ In addition, recent studies have identified several GOT1 inhibitors^{58–60} that may provide new ideas for potential strategies to target GOT2. As a KAT2 inhibitor, PF-04859989 inhibits GOT1 activity in a time- and pyridoxal-5'-phosphate (PLP)-dependent manner and selectively impairs pancreatic cancer cell line growth. Unfortunately, the inhibitory activity of PF-04859989 on GOT2 is relatively low.⁵⁸ Therefore, the development of potent GOT2 inhibitors is a challenging next step.

Before these findings can be applied to the clinic, some important issues remain to be resolved. First, the selection of preclinical models may cause differences in study results. Kerk et al¹⁹ and Garcia-Bermudez et al²¹ found that proliferation of pancreatic cancer cells *in vitro* is dependent on GOT2, but that GOT2 is not required for tumor growth in immunodeficient or immunocompetent mouse xenograft models of pancreatic cancer. However, Abrego et al obtained the exact opposite results.⁸ GOT2 knockout did not affect the proliferation of pancreatic cancer cells *in vitro* and tumor progression in immunocompromised mouse models, but severely impaired xenograft growth in mice with intact immune systems. This prompts us to develop mouse models that more accurately reproduce human disease to fully understand how the metabolic and immune effects of GOT2 interact in pancreatic cancer. Secondly, targeting the GOT2-PPAR δ axis may have great therapeutic potential. This requires us to further clarify the regulatory mechanisms of GOT2 nuclear localization and the precise molecular mechanisms by which GOT2 regulates PPAR δ transcriptional activity. Moreover, recent studies have revealed compensatory mechanisms of pancreatic cancer to bypass GOT2 dependence,^{19,21} which prompts us to target resistance mechanisms for improved treatment and to focus on whether fibroblasts mediate resistance to GOT2 silencing in the intact immune system. Finally, because pancreatic cancer patients suffer from severe immunosuppression and dysfunction of T-cell populations,⁶¹ whether targeting GOT2 is sufficient to reverse the immune suppression in this case remains to be investigated.

Author contributions

J.B.: writing – original draft, visualization. Z.M.: writing – original draft, conceptualization. Q.Y.: writing – review & editing, supervision. All authors read and approved the final version of this manuscript.

Conflict of interests

The authors declared no competing interests.

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References

- Ullman NA, Burchard PR, Dunne RF, Linehan DC. Immunologic strategies in pancreatic cancer: making cold tumors Hot. *J Clin Oncol.* 2022;40(24):2789–2805.
- Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrias LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res.* 2014;74(11):2913–2921.
- Vincent A, Herman J, Schulick R, Hruban RH, Goggins M. Pancreatic cancer. *Lancet.* 2011;378(9791):607–620.
- Birsoy K, Wang T, Chen WW, Freinkman E, Abu-Remaileh M, Sabatini DM. An essential role of the mitochondrial electron transport chain in cell proliferation is to enable aspartate synthesis. *Cell.* 2015;162(3):540–551.
- Sullivan LB, Gui DY, Hosios AM, Bush LN, Freinkman E, Vander Heiden MG. Supporting aspartate biosynthesis is an essential function of respiration in proliferating cells. *Cell.* 2015;162(3):552–563.
- Xiao D, Zeng L, Yao K, Kong X, Wu G, Yin Y. The glutamine-alpha-ketoglutarate (AKG) metabolism and its nutritional implications. *Amino Acids.* 2016;48(9):2067–2080.
- Borst P. The malate-aspartate shuttle (Borst cycle): how it started and developed into a major metabolic pathway. *IUBMB Life.* 2020;72(11):2241–2259.
- Abrego J, Sanford-Crane H, Oon C, et al. A cancer cell-intrinsic GOT2-PPAR δ axis suppresses antitumor immunity. *Cancer Discov.* 2022;12(10):2414–2433.
- Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metabol.* 2016;23(1):27–47.
- Martínez-Reyes I, Chandel NS. Cancer metabolism: looking forward. *Nat Rev Cancer.* 2021;21(10):669–680.
- Vander Heiden MG. Targeting cancer metabolism: a therapeutic window opens. *Nat Rev Drug Discov.* 2011;10(9):671–684.
- Son J, Lyssiotis CA, Ying H, et al. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature.* 2013;496(7443):101–105.
- Chakrabarti G, Moore ZR, Luo X, et al. Targeting glutamine metabolism sensitizes pancreatic cancer to PARP-driven metabolic catastrophe induced by β -lapachone. *Cancer Metabol.* 2015;3:12.
- Bryant KL, Mancias JD, Kimmelman AC, Der CJ. KRAS: feeding pancreatic cancer proliferation. *Trends Biochem Sci.* 2014;39(2):91–100.
- Ying H, Kimmelman AC, Lyssiotis CA, et al. Oncogenic Kras maintains pancreatic tumors through regulation of anabolic glucose metabolism. *Cell.* 2012;149(3):656–670.
- Wise DR, Thompson CB. Glutamine addiction: a new therapeutic target in cancer. *Trends Biochem Sci.* 2010;35(8):427–433.
- DeBerardinis RJ, Cheng T. Q's next: the diverse functions of glutamine in metabolism, cell biology and cancer. *Oncogene.* 2010;29(3):313–324.
- Shanware NP, Mullen AR, DeBerardinis RJ, Abraham RT. Glutamine: pleiotropic roles in tumor growth and stress resistance. *J Mol Med.* 2011;89(3):229–236.
- Kerk SA, Lin L, Myers AL, et al. Metabolic requirement for GOT2 in pancreatic cancer depends on environmental context. *Elife.* 2022;11:e73245.
- Yang S, Hwang S, Kim M, Seo SB, Lee JH, Jeong SM. Mitochondrial glutamine metabolism via GOT2 supports pancreatic

- cancer growth through senescence inhibition. *Cell Death Dis.* 2018;9(2):55.
- 21. Garcia-Bermudez J, Badgley MA, Prasad S, et al. Adaptive stimulation of macropinocytosis overcomes aspartate limitation in cancer cells under hypoxia. *Nat Metab.* 2022;4(6):724–738.
 - 22. Collado M, Serrano M. Senescence in tumours: evidence from mice and humans. *Nat Rev Cancer.* 2010;10(1):51–57.
 - 23. Majumder PK, Grisanzio C, O’Connell F, et al. A prostatic intraepithelial neoplasia-dependent p27 Kip1 checkpoint induces senescence and inhibits cell proliferation and cancer progression. *Cancer Cell.* 2008;14(2):146–155.
 - 24. Jones S, Zhang X, Parsons DW, et al. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. *Science.* 2008;321(5897):1801–1806.
 - 25. Yang H, Zhou L, Shi Q, et al. SIRT 3-dependent GOT 2 acetylation status affects the malate–aspartate NADH shuttle activity and pancreatic tumor growth. *EMBO J.* 2015;34(8):1110–1125.
 - 26. Chen Y, Fu LL, Wen X, et al. Sirtuin-3 (SIRT3), a therapeutic target with oncogenic and tumor-suppressive function in cancer. *Cell Death Dis.* 2014;5(2):e1047.
 - 27. Janes MR, Zhang J, Li LS, et al. Targeting KRAS mutant cancers with a covalent G¹²C-specific inhibitor. *Cell.* 2018;172(3):578–589.e17.
 - 28. Jänne PA, Riely GJ, Gadgeel SM, et al. Adagrasib in non-small-cell lung cancer harboring a KRASG12C mutation. *N Engl J Med.* 2022;387(2):120–131.
 - 29. Jin H, Wang S, Zaal EA, et al. A powerful drug combination strategy targeting glutamine addiction for the treatment of human liver cancer. *Elife.* 2020;9:e56749.
 - 30. Gross MI, Demo SD, Dennison JB, et al. Antitumor activity of the glutaminase inhibitor CB-839 in triple-negative breast cancer. *Mol Cancer Therapeut.* 2014;13(4):890–901.
 - 31. Hong R, Zhang W, Xia X, et al. Preventing BRCA1/ZBRK1 repressor complex binding to the GOT2 promoter results in accelerated aspartate biosynthesis and promotion of cell proliferation. *Mol Oncol.* 2019;13(4):959–977.
 - 32. Jin M, Shi C, Hua Q, et al. High circ-SEC31A expression predicts unfavorable prognoses in non-small cell lung cancer by regulating the miR-520a-5p/GOT-2 axis. *Aging.* 2020;12(11):10381–10397.
 - 33. Li Y, Li B, Xu Y, et al. GOT2 silencing promotes reprogramming of glutamine metabolism and sensitizes hepatocellular carcinoma to glutaminase inhibitors. *Cancer Res.* 2022;82(18):3223–3235.
 - 34. Meléndez-Rodríguez F, Urrutia AA, Lorendeau D, et al. HIF1 α suppresses tumor cell proliferation through inhibition of aspartate biosynthesis. *Cell Rep.* 2019;26(9):2257–2265.e4.
 - 35. Du F, Chen J, Liu H, et al. SOX12 promotes colorectal cancer cell proliferation and metastasis by regulating asparagine synthesis. *Cell Death Dis.* 2019;10(3):239.
 - 36. Adhikary T, Kadatz K, Finkernagel F, et al. Genomewide analyses define different modes of transcriptional regulation by peroxisome proliferator-activated receptor- β/δ (PPAR β/δ). *PLoS One.* 2011;6(1):e16344.
 - 37. Markosyan N, Li J, Sun YH, et al. Tumor cell-intrinsic EPHA2 suppresses anti-tumor immunity by regulating PTGS2 (COX-2). *J Clin Investig.* 2019;129(9):3594–3609.
 - 38. Zelenay S, van der Veen AG, Böttcher JP, et al. Cyclooxygenase-dependent tumor growth through evasion of immunity. *Cell.* 2015;162(6):1257–1270.
 - 39. Hickman TL, Choi E, Whiteman KR, et al. BOXR1030, an anti-GPC3 CAR with exogenous GOT2 expression, shows enhanced T cell metabolism and improved anti-cell line derived tumor xenograft activity. *PLoS One.* 2022;17(5):e0266980.
 - 40. Feist M, Schwarzfischer P, Heinrich P, et al. Cooperative STAT/NF- κ B signaling regulates lymphoma metabolic reprogramming and aberrant GOT2 expression. *Nat Commun.* 2018;9:1514.
 - 41. Yang R, Cheng S, Xiao J, et al. GLS and GOT2 as prognostic biomarkers associated with dendritic cell and immunotherapy response in breast cancer. *Heliyon.* 2024;10(1):e24163.
 - 42. Zhang Y, Crawford HC, Pasca di Maglino M. Epithelial-stromal interactions in pancreatic cancer. *Annu Rev Physiol.* 2019;81:211–233.
 - 43. Storz P, Crawford HC. Carcinogenesis of pancreatic ductal adenocarcinoma. *Gastroenterology.* 2020;158(8):2072–2081.
 - 44. Koong AC, Mehta VK, Le QT, et al. Pancreatic tumors show high levels of hypoxia. *Int J Radiat Oncol.* 2000;48(4):919–922.
 - 45. Garcia-Bermudez J, Baudrier L, La K, et al. Aspartate is a limiting metabolite for cancer cell proliferation under hypoxia and in tumours. *Nat Cell Biol.* 2018;20(7):775–781.
 - 46. Zhao H, Yang L, Baddour J, et al. Tumor microenvironment derived exosomes pleiotropically modulate cancer cell metabolism. *Elife.* 2016;5:e10250.
 - 47. Commissio C, Davidson SM, Soydaner-Azeloglu RG, et al. Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature.* 2013;497(7451):633–637.
 - 48. Davidson SM, Jonas O, Keibler MA, et al. Direct evidence for cancer-cell-autonomous extracellular protein catabolism in pancreatic tumors. *Nat Med.* 2017;23(2):235–241.
 - 49. Kamphorst JJ, Nofal M, Commissio C, et al. Human pancreatic cancer tumors are nutrient poor and tumor cells actively scavenge extracellular protein. *Cancer Res.* 2015;75(3):544–553.
 - 50. Wykoff CC, Beasley NJ, Watson PH, et al. Hypoxia-inducible expression of tumor-associated carbonic anhydrases. *Cancer Res.* 2000;60(24):7075–7083.
 - 51. Boone BA, Bahary N, Zureikat AH, et al. Safety and biologic response of pre-operative autophagy inhibition in combination with gemcitabine in patients with pancreatic adenocarcinoma. *Ann Surg Oncol.* 2015;22(13):4402–4410.
 - 52. Zeh HJ, Bahary N, Boone BA, et al. A randomized phase II pre-operative study of autophagy inhibition with high-dose hydroxychloroquine and gemcitabine/nab-paclitaxel in pancreatic cancer patients. *Clin Cancer Res.* 2020;26(13):3126–3134.
 - 53. Supuran CT, Winum JY. Carbonic anhydrase IX inhibitors in cancer therapy: an update. *Future Med Chem.* 2015;7(11):1407–1414.
 - 54. Liu J, Xia H, Kim M, et al. Beclin1 controls the levels of p53 by regulating the deubiquitination activity of USP10 and USP13. *Cell.* 2011;147(1):223–234.
 - 55. Kulkarni YM, Kaushik V, Azad N, et al. Autophagy-induced apoptosis in lung cancer cells by a novel digitoxin analog. *J Cell Physiol.* 2016;231(4):817–828.
 - 56. Thornburg JM, Nelson KK, Clem BF, et al. Targeting aspartate aminotransferase in breast cancer. *Breast Cancer Res.* 2008;10(5):R84.
 - 57. Wise DR, DeBerardinis RJ, Mancuso A, et al. Myc regulates a transcriptional program that stimulates mitochondrial glutaminolysis and leads to glutamine addiction. *Proc Natl Acad Sci U S A.* 2008;105(48):18782–18787.
 - 58. Yoshida T, Yamasaki S, Kaneko O, et al. A covalent small molecule inhibitor of glutamate-oxaloacetate transaminase 1 impairs pancreatic cancer growth. *Biochem Biophys Res Commun.* 2020;522(3):633–638.
 - 59. Sun W, Luan S, Qi C, et al. Aspulvinone O, a natural inhibitor of GOT1 suppresses pancreatic ductal adenocarcinoma cells growth by interfering glutamine metabolism. *Cell Commun Signal.* 2019;17(1):111.
 - 60. Anglin J, Zavareh RB, Sander PN, et al. Discovery and optimization of aspartate aminotransferase 1 inhibitors to target redox balance in pancreatic ductal adenocarcinoma. *Bioorg Med Chem Lett.* 2018;28(16):2675–2678.
 - 61. Steele NG, Carpenter ES, Kemp SB, et al. Multimodal mapping of the tumor and peripheral blood immune landscape in human pancreatic cancer. *Nat Can (Ott).* 2020;1(11):1097–1112.